

RxODE and **nlmixr**: open-source packages for pharmacometric modelling in R

Pharmacometrics Network Benelux Presentation

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RxODE is pharmacometric simulation software as an open-source R package

- Written by Wenping Wang and Matt Fidler, available on CRAN¹ and GitHub², and described in a tutorial in CPT:PSP³
- Simulation of ODEs was already possible in R (using deSolve), but was slow and virtually impossible to code with flexible dosing history
- RxODE has rapid execution due to compilation in C
- RxODE allows fully flexible dosing history
- Stable and mature software for Windows, OS X, Linux
- Requires external compilers (provided by Rtools on Windows)

- New developments (alpha stage): parallelisation to increase speed even further

[1] CRAN: <https://cran.r-project.org/web/packages/RxODE/index.html>

[2] GitHub: <https://github.com/nlmixrdevelopment/RxODE>

[3] Wang W et al. CPT:PSP (2016) 5, 3–10.

Basic example

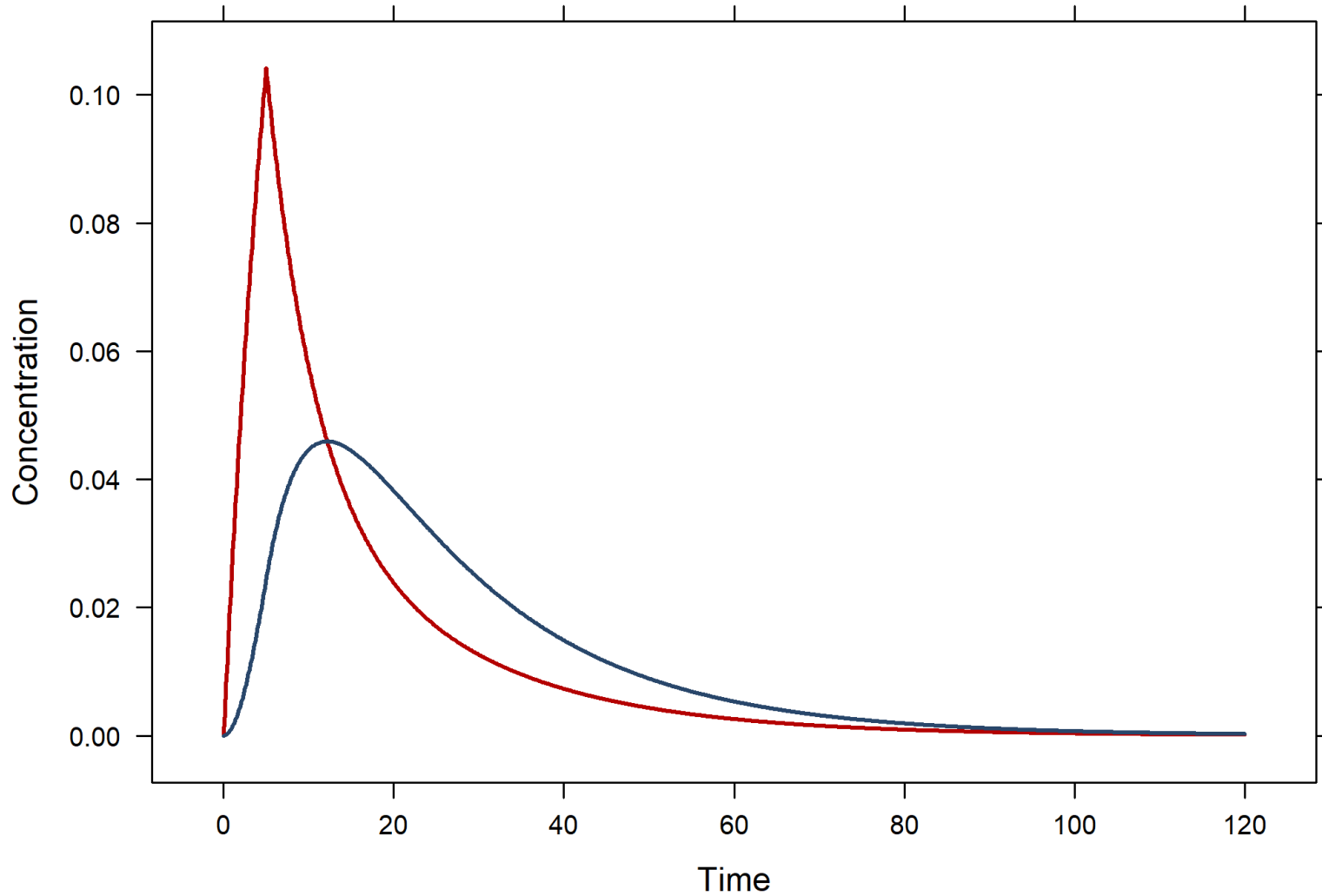
```
library(RxODE)

ode1 <- "
  K12 = CL2/V;
  K21 = CL2/V2;
  d/dt(centr) = K21*periph-K12*centr-(VMAX*centr/V) / (KM+centr/V) -CL*centr/V;
  d/dt(periph) =-K21*periph+K12*centr;
  C1=centr/V;
  C2=periph/V2;
"
mod1 <- RxODE(model = ode1, modName = 'mod1')

ev <- eventTable()
ev$add.dosing(
  dose = 10,
  nbr.doses = 1,
  dosing.to = 1,
  rate = 2,
  start.time = 0
)
ev$add.sampling(seq(0,120,0.1))
Params <- c(VMAX=2000, KM=700, CL=4, CL2=3, V=70, V2=30)
Res <- as.data.frame(mod1$run(Params, ev))

xyplot(C1+C2~time, data=Res, type='l', ylab="Concentration", xlab="Time")
```

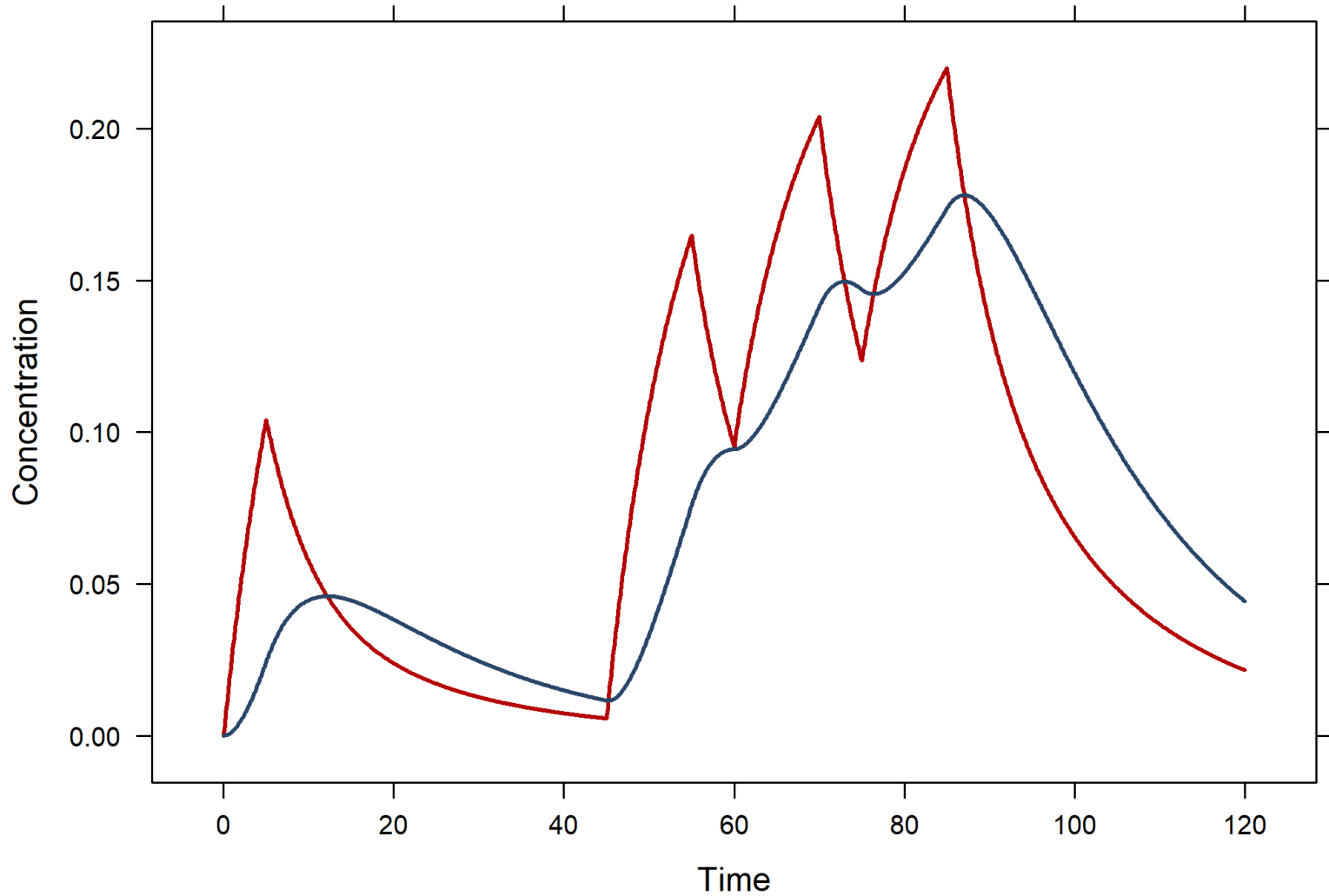
Single dose



Adding extra doses (expand the event table)

```
ev$add.dosing(  
  dose = 20,  
  nbr.doses = 3,  
  dosing.to = 1,  
  dosing.interval=15,  
  rate = 2,  
  start.time = 45  
)  
res<-as.data.frame(mod1$run(Params, ev))  
  
xyplot(C1+C2~time, data=res, type='l', ylab="Concentration", xlab="Time")
```

Multiple dose



Generate a whole population of full individual IPRED curves starting from a NONMEM dataset (study this at home 😊)

```
library(data.table)
NMdat <- fread(file.path(datapath, "run100.csv"))
EBEs <- unique(NMdat[, .(ID, STD, VMAX, KM, CL, CL2, V, V2)])
subs <- unique(NMdat$ID)
N <- length(subs)
s = lapply(1:N, function(i) {
  params <- EBEs[ID == subs[i]]
  ev <- eventTable()
  DOSi <- NMdat[ID == subs[i] & AMT > 0]
  DOSi[, nTime := shift(TIME, 1L, type = 'lead')]
  timei <- NMdat$TIME[NMdat$ID == subs[i]]
  for (j in 1:length(DOSi$AMT)) {
    dos <- DOSi[j, ]
    ev$add.dosing(dose = dos$AMT, nbr.doses = 1, dosing.to = 1,
                 rate = dos$RATE, start.time = dos$TIME)
    #generate prediction time points (many points at dose and fewer at later times)
    if (is.na(dos$nTime)) {dos$nTime <- dos$TIME + 720}
    timei <- c(timei, dos$TIME + exp(seq(log(+0.01), log(dos$nTime - dos$TIME - 0.01),
                                       (log(dos$nTime - dos$TIME - 0.01) - log(+0.01)) / 100)))
  }
  times <- sort(unique(timei))
  ev$add.sampling(times)
  x <- as.data.table(mod1$run(params, ev))
  x[, ID := subs[i]]
  setnames(x, "C1", "IPRED")
})
df.sim = as.data.table(do.call("rbind", s))
```

You need to simulate before you can estimate

- With simulation covered, you can start to think about estimation
- Combine the simulation core with estimation routines and you get:

nlmixr!

nlmixr is an open-source R package

- Written by Wenping Wang and Matt Fidler, and available on GitHub and CRAN^{1,2}:
 - builds on RxODE³
 - combined with nlme and SAEM estimation routines, provides an R package for parameter estimation in nonlinear mixed effect models
 - much, more to come (e.g. adaptive Gaussian quadrature for non-continuous data, and with FOCE-I under development)
- **nlmixr** is completely free and open, and does not depend on any other commercial tool such as NONMEM or Monolix
- **nlmixr** provides an efficient and versatile way to specify pharmacometric models (both closed-form and ODEs) and dosing scenarios, with rapid execution due to compilation in C

[1] <https://github.com/nlmixrdevelopment/nlmixr>

[2] <https://cran.r-project.org/web/packages/RxODE/index.html>

[3] Wang W et al. CPT:PSP (2016) 5, 3–10.

nlmixr is an open-source R package

- Models are defined using a unified user interface (UI): common input and output structure for the various estimation algorithms
- `xpose.nlmixr`¹ written by Justin Wilkins provides linkage to the new Xpose package², written by Ben Guiaستنec, feeding the uniform output into a highly flexible diagnostics package
- The `shinyMixR`³ project management tool written by Richard Hooijmaijers and Teun Post provides an interface to `nlmixr` from both the R command line and a user-friendly browser-based Shiny dashboard application
- `nlmixr` requires access to compilers (e.g. using Rtools) and Python: both a full-package windows installer is available, and instructions on managing your own installation
- Documentation is available in the form of a bookdown (nlmixr.github.io) written and curated by Teun Post
- Runs on Linux, Windows, and OS X

[1] <https://github.com/nlmixrdevelopment/xpose.nlmixr>

[2] <https://CRAN.R-project.org/package=xpose>

[3] <https://github.com/RichardHooijmaijers/shinyMixR>

The unified user interface

- Models are defined using a function containing an initialisation block (`ini`) and a model definition block (`model`)

```
mod1 <- function() {  
  ini({  
  
  })  
  model({  
  
  })  
}
```

The unified user interface

- The `ini` block defines the parameters
 - Thetas defined using assign operators (`<-` or `=`)
 - Residual error defined using assign operators (`<-` or `=`)
 - Etas defined using a model formula (`~`)
- Parameter names, starting values, labels (using `#`), bounds

```
mod1 <- function() {  
  ini({  
    lCl <- 1.6      #log Cl (L/hr)  
    lVc = log(90)  #log V (L)  
    lKa <- 1       #log Ka (1/hr)  
    prop.err <- c(0, 0.2, 1)  
    eta.Ka ~ 0.1   #IIV Ka  
    eta.Cl + eta.Vc ~ c(0.1,  
                        0.005, 0.1)  
  })  
  model({  
  })  
}
```

The unified user interface

- The `model` block defines
 - the relationship between thetas and etas
 - the model structure using either ODEs or closed-form solutions
 - the residual error structure and where it is applied

```
mod1 <- function() {  
  ini({  
  
  })  
  model({  
    Cl <- exp(lCl + eta.Cl)  
    Vc <- exp(lVc + eta.Vc)  
    KA <- exp(lKa + eta.Ka)  
    kel <- Cl / Vc  
    d/dt(depot) = -KA*depot  
    d/dt(centr) = KA*depot - kel*centr  
    cp = centr / Vc  
    cp ~ prop(prop.err)  
  })  
}
```

Parameterisation and mu-referencing

- For SAEM, parameters must be defined using 'mu-referencing' and this implies estimating log-parameters with the IIV added on the log-scale
- For nlme, mu-referencing is not strictly required, but is shown to provide superior estimation results

```
Data$logWT70 <- log(Data$WT/70)
```

```
mod1 <- function(){  
  ini({  
    ## For SAEM parameters must be defined using 'mu-referencing'  
    ## For nlme mu-referencing is not strictly required but is shown to provide  
    ## superior estimation results  
    lCl <- 1.6      #log Cl (L/hr)  
    AllomCl <- 0.75 #log Cl (L/hr)  
    ## .....  
    eta.Cl ~ 0.1   # IIV Cl  
    ## .....  
  })  
  model({  
    ## Parameters are defined in terms of the initial estimates  
    Cl <- exp(lCl + eta.Cl)  
    ## or for implementing covariate effects:  
    Cl <- exp(lCl + eta.Cl + logWT70*AllomCl)  
    ## Data transformations should be done outside the model definition  
    ## .....  
  })  
}
```

Full example with proportional and additive error

```
mod1 <- function() {
  ini({
    ## Initial conditions for population parameters (sometimes
    ## called theta parameters) are defined by either `<-` or `=`
    lCl <- 1.6      #log Cl (L/hr)
    ## Note that simple expressions that evaluate to a number are
    ## OK for defining initial conditions (like in R)
    lVc = log(90)  #log V (L)
    ## Also a comment on a parameter is captured as a parameter label
    lKA <- 0.1     #log Ka (1/hr)
    ## Bounds may be specified by c(lower, est, upper), like NONMEM:
    ## Residuals errors are assumed to be population parameters
    prop.err <- c(0, 0.2, 1)
    add.err  <- c(0,0.01)
    ## Initial estimate for ka IIV variance
    ## Labels work for single parameters
    eta.Cl ~ 0.1   # IIV Cl
    ## For correlated parameters, you specify the names of each
    ## correlated parameter separated by a addition operator `+`
    ## and the left handed side specifies the lower triangular
    ## matrix initial of the covariance matrix.
    eta.Vc + eta.KA ~ c(0.1,
                       0.005, 0.1)

    ## Note that labels are not defined for correlated parameters.
  })
  model({
    ## Parameters are defined in terms of the initial estimates
    Cl <- exp(lCl + eta.Cl)
    Vc <- exp(lVc + eta.Vc)
    KA <- exp(lKA + eta.KA)
    ## Next, the differential equations are defined
    kel <- Cl / Vc;
    d/dt(depot) = -KA*depot;
    d/dt(centr) = KA*depot-kel*centr;
    ## And the concentration is then calculated
    cp = centr / Vc;
    ## Last, nlmixr is told that the plasma concentration follows
    ## a combined proportional/additive error
    cp ~ prop(prop.err)+add(add.err)
  })
}
```

And using a closed-form solution

```
mod1 <- function(){
  ini({
    lCl <- 1.6      #log Cl (L/hr)
    lVc <- 4.5      #log V (L)
    lKA <- 0.1      #log Ka (1/hr)
    prop.err <- c(0, 0.2, 1)
    eta.Cl ~ 0.1    # IIV Cl
    eta.Vc + eta.KA ~ c(0.1,
                        0.005, 0.1)
  })
  model({
    Cl <- exp(lCl + eta.Cl)
    Vc <- exp(lVc + eta.Vc)
    KA <- exp(lKA + eta.KA)
    ## Instead of specifying the ODEs, you can use
    ## the linCmt() function to use closed-form solutions.
    ## This function determines the type of PK solved system
    ## to use by the parameters that are defined.
    ## In this case it knows that this is a one-compartment model
    ## with first-order absorption.
    linCmt() ~ prop(prop.err)
  })
}
```


The nlmixr dataset

- Datasets need to comply with RxODE requirements
- EVID is more complex
 - 101 for bolus dose in compartment 1, 10101 for infusion in compartment 1
- No MDV item so no on-the-fly removal of unwanted records
- Infusions need two records: one to start infusion and one to stop infusion (at time of infusion stop with a negative rate)
- No SS dosing so steady state needs to be coded using multiple preceding doses
- NONMEM datasets can be converted using a special function based on code by Yuan Xiong:

```
dat <- nmDataConvert (dat);
```

Running nlmixr

- `nlmixr` is run using the following structure:

```
fit <- nlmixr(model.function,  
             rxode.dataset,  
             est = "est",  
             control = estControl(options))
```

- Currently `nlme` and `SAEM` are implemented

- Example for `nlme`:

```
fit <- nlmixr(mod1,  
             dat,  
             est = "nlme",  
             control = nlmeControl(pnlstol = .05))
```

- Example for `SAEM`:

```
fit <- nlmixr(mod1,  
             dat,  
             est = "saem",  
             control = saemControl(  
               n.burn = 200,  
               n.em = 300,  
               print = 50  
             ))
```

The shinyMixR interface can manage your runs

The screenshot displays the RStudio interface for the shinyMixR package. The main editor shows an R script with the following code:

```
1 # Example of workflow directly within R
2 library(shinyMixR)
3
4 # Create new project
5 create_proj()
6
7 # Obtain project information
8 proj <- get_proj()
9
10 # Run nlmixr models (async in separate Rsession)
11 run_nmx("run1",proj)
12
13 # Use results created by the package
14 res <- readRDS("shinyMixR/run1.res.rds")
15 ggplot(res,aes(DV,PRED)) + geom_point(alpha=.6) + geom_abline(intercept=0,slope=1,colour="darkblue",linetype=2)
16
17 # Several other functions written for the interface are available for use in R environment
18 oview() # Create data frame with overview of the models within the project
19 makeTree() # Create interactive tree view of the models within project
20
```

The Environment pane on the right shows the following data and functions:

Global Environment	
Data	
res	132 obs. of 15 variables
Functions	
AdaptModel	function (projlst, inp, session)
AdaptOverview	function (projlst, inp, session, type = "modal")
makeTree	function ()
oview	function ()
oviewUI	function (projlst)

The Workflow Viewer pane on the right shows a diagram of the execution flow:

```
graph LR
  start((start)) --> run1((run1))
  start --> run2((run2))
  start --> run7((run7))
  run1 --> run3((run3))
  run2 --> run4((run4))
  run2 --> run6((run6))
  run4 --> run5((run5))
```

The shinyMixR interface can be run from R...

```
# Example of workflow directly within R
library(shinyMixR)

# Create new project
create_proj()

# Obtain project information
proj <- get_proj()

# Run nlmixr models (async in separate Rsession)
run_nmx("run1",proj)

# Use results created by the package
res <- readRDS("shinyMixR/run1.res.rds")
ggplot(res,aes(DV,PRED)) + geom_point(alpha=.6) + geom_abline(intercept=0,slope=1,colour="darkblue",linetype=2)

# Several other functions written for the interface are available for use in R environment
oview() # Create data frame with overview of the models within the project
makeTree() # Create interactive tree view of the models within project
```

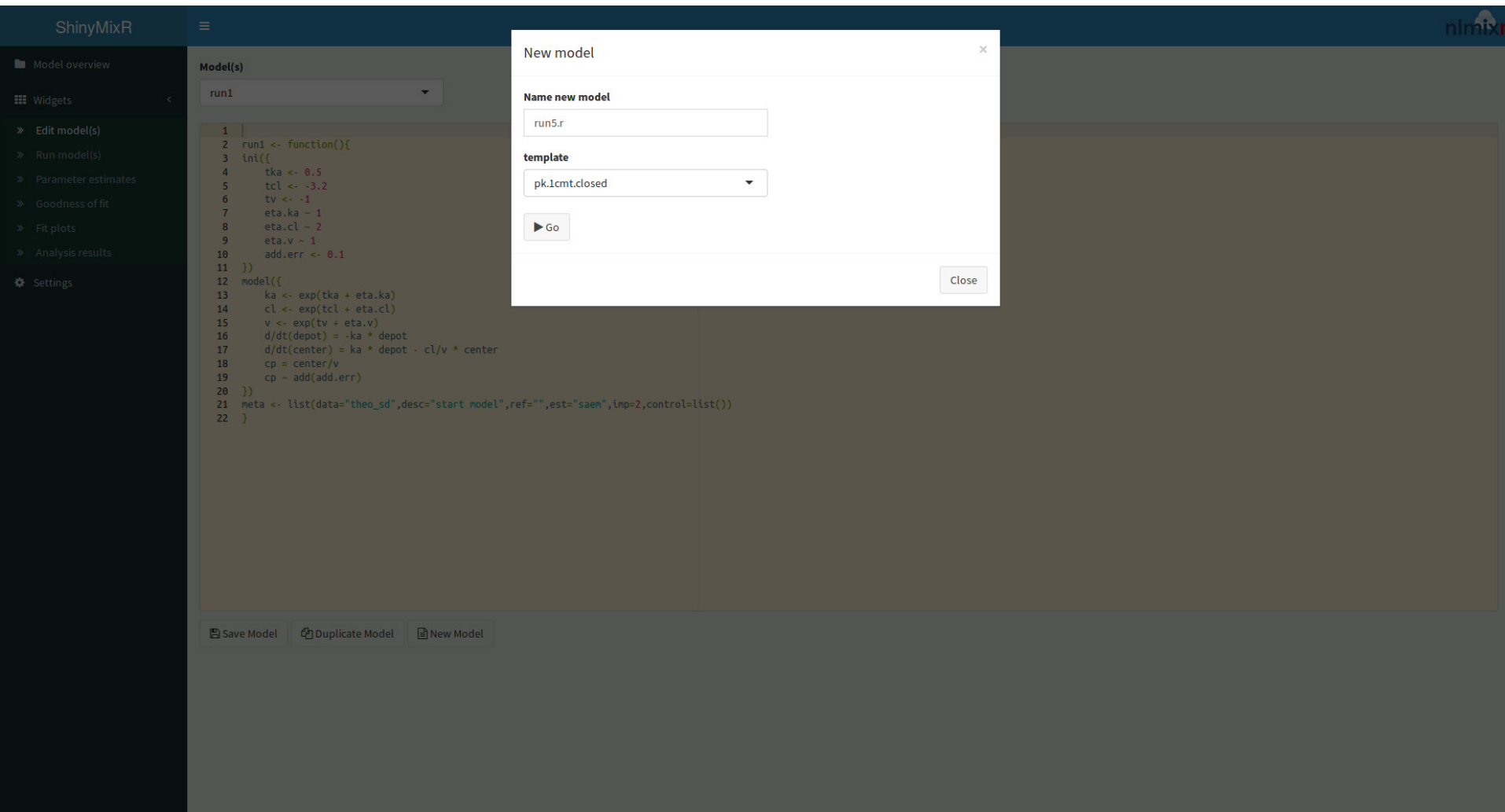
...or by launching a browser session

The screenshot displays the ShinyMixR web interface. On the left is a dark sidebar with navigation options: Model overview, Widgets, Edit model(s), Run model(s), Parameter estimates, Goodness of fit, Fit plots, Analysis results, and Settings. The main content area has a blue header with 'ShinyMixR' and a hamburger menu icon. Below the header are 'Refresh' and 'Adapt model notes' buttons. The 'Overview' section features a table with columns: models, importance, description, ref, data, method, OBJF, dOBJF, and runtime. Below the table are filter buttons for each column, all set to 'All'. The 'Tree View' section contains a 'make tree' button and a graph showing a flow from 'start' to 'run1', which then branches into 'run2' and 'run3', with 'run2' further leading to 'run4'.

models	importance	description	ref	data	method	OBJF	dOBJF	runtime
run1	2	start model		theo_sd	saem	116.137		31.827
run2	3	test one	run1	theo_md	saem	349.627	233.49	79.909
run3	2	test two	run1	theo_sd	saem	116.137	0	24.843
run4	0	test three	run2		saem			

```
graph LR; start((start)) --> run1((run1)); run1 --> run2((run2)); run1 --> run3((run3)); run2 --> run4((run4))
```

Where models can be edited and run...



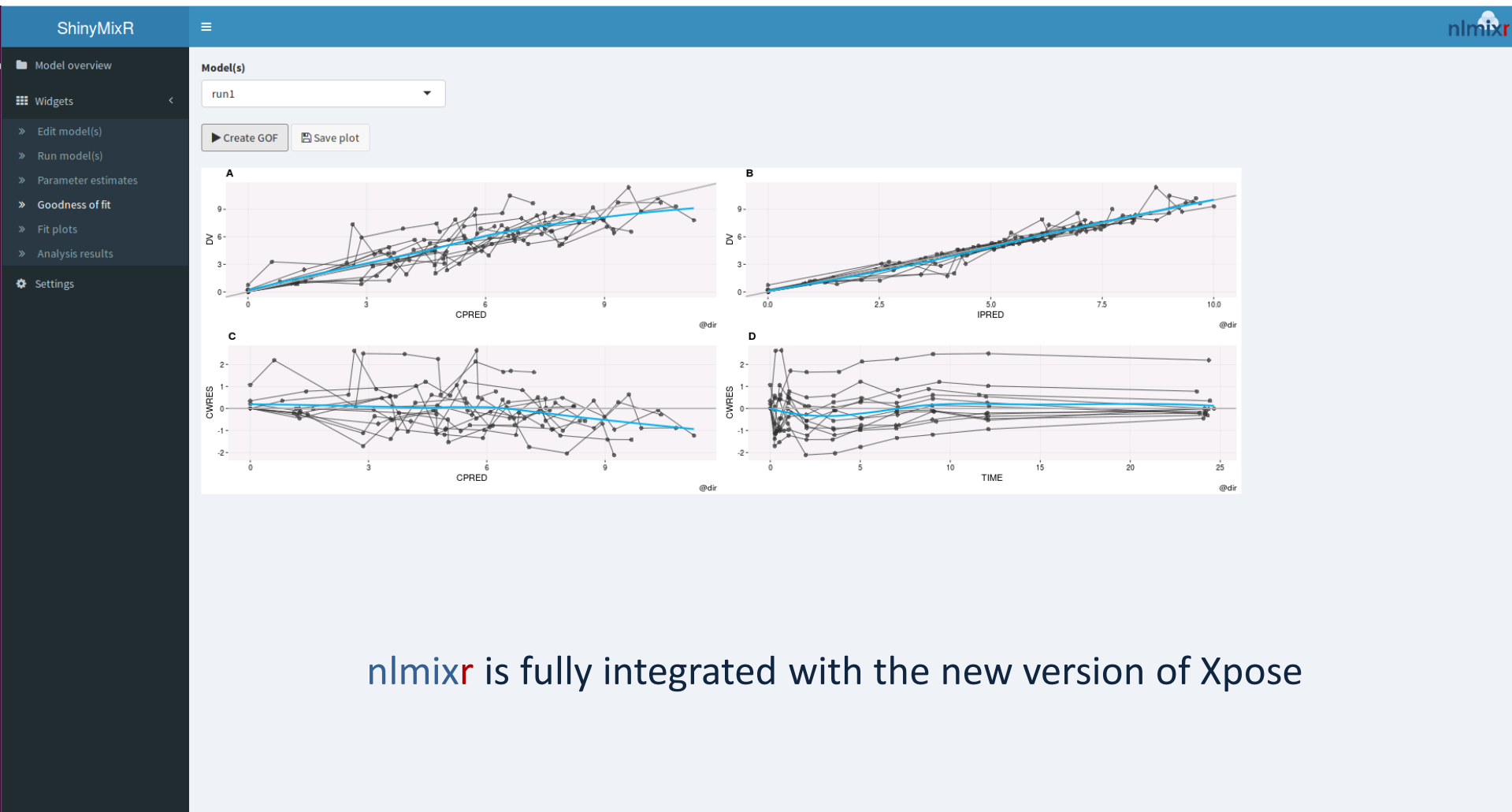
The screenshot displays the ShinyMixR web application interface. On the left, a dark sidebar contains navigation options: 'Model overview', 'Widgets', 'Edit model(s)', 'Run model(s)', 'Parameter estimates', 'Goodness of fit', 'Fit plots', 'Analysis results', and 'Settings'. The main area shows a code editor with R code for a model named 'run1'. The code includes initial values for parameters (tka, tcl, tv, eta.ka, eta.cl, eta.v, add.err) and a model definition with differential equations for depot and center concentrations, along with a meta-data list.

A 'New model' dialog box is open in the foreground. It has a title bar with a close button (X). The dialog contains the following fields and controls:

- Name new model:** A text input field containing 'run5.r'.
- template:** A dropdown menu with 'pk.1cmt.closed' selected.
- Go:** A button with a right-pointing triangle icon.
- Close:** A button located at the bottom right of the dialog.

At the bottom of the main interface, there are three buttons: 'Save Model', 'Duplicate Model', and 'New Model'.

...and output like goodness of fit plots can be created using the new Xpose functionality, or using custom scripts...



The screenshot displays the ShinyMixR web application interface. On the left is a dark sidebar with navigation options: Model overview, Widgets, Edit model(s), Run model(s), Parameter estimates, Goodness of fit, Fit plots, Analysis results, and Settings. The main content area shows a 'Model(s)' dropdown set to 'run1' and buttons for 'Create GOF' and 'Save plot'. Below this are four plots labeled A, B, C, and D. Plot A shows DV vs CPRED, Plot B shows DV vs IPRED, Plot C shows CWRES vs CPRED, and Plot D shows CWRES vs TIME. Each plot contains multiple grey lines representing individual data points and a solid blue line representing the model fit. The plots are arranged in a 2x2 grid.

nlmixr is fully integrated with the new version of Xpose

...and individual plots as well

The screenshot displays the ShinyMixR web application interface. On the left, a dark sidebar contains navigation options: Model overview, Widgets, Edit model(s), Run model(s), Parameter estimates, Goodness of fit, Fit plots, Analysis results, and Settings. The main content area shows a 'Model(s)' dropdown set to 'run1' and buttons for 'Create Fit plots' and 'Save plot'. Below these are three vertically stacked plot windows, each showing 'DV' vs 'TIME' for a different model instance (1, 5, and 9). Each plot features observed data points (black dots), a solid line representing the model fit, and a dashed line representing the confidence interval. The y-axis ranges from 0 to 9, and the x-axis ranges from 0 to 25. A window titled 'IndFit.pdf' is overlaid on the right, displaying a grid of 12 individual plots (labeled 1 through 12) in a 3x4 arrangement. The window title bar shows 'IndFit.pdf', '1 of 1', navigation icons, a zoom level of '78,49%', and a '1 of 1' page indicator. The date 'January 12, 2018' is visible in the top left of the PDF window. The caption 'Figure 1: plot' is centered above the grid. Each plot in the grid shows the same 'DV' vs 'TIME' data as the main application window.

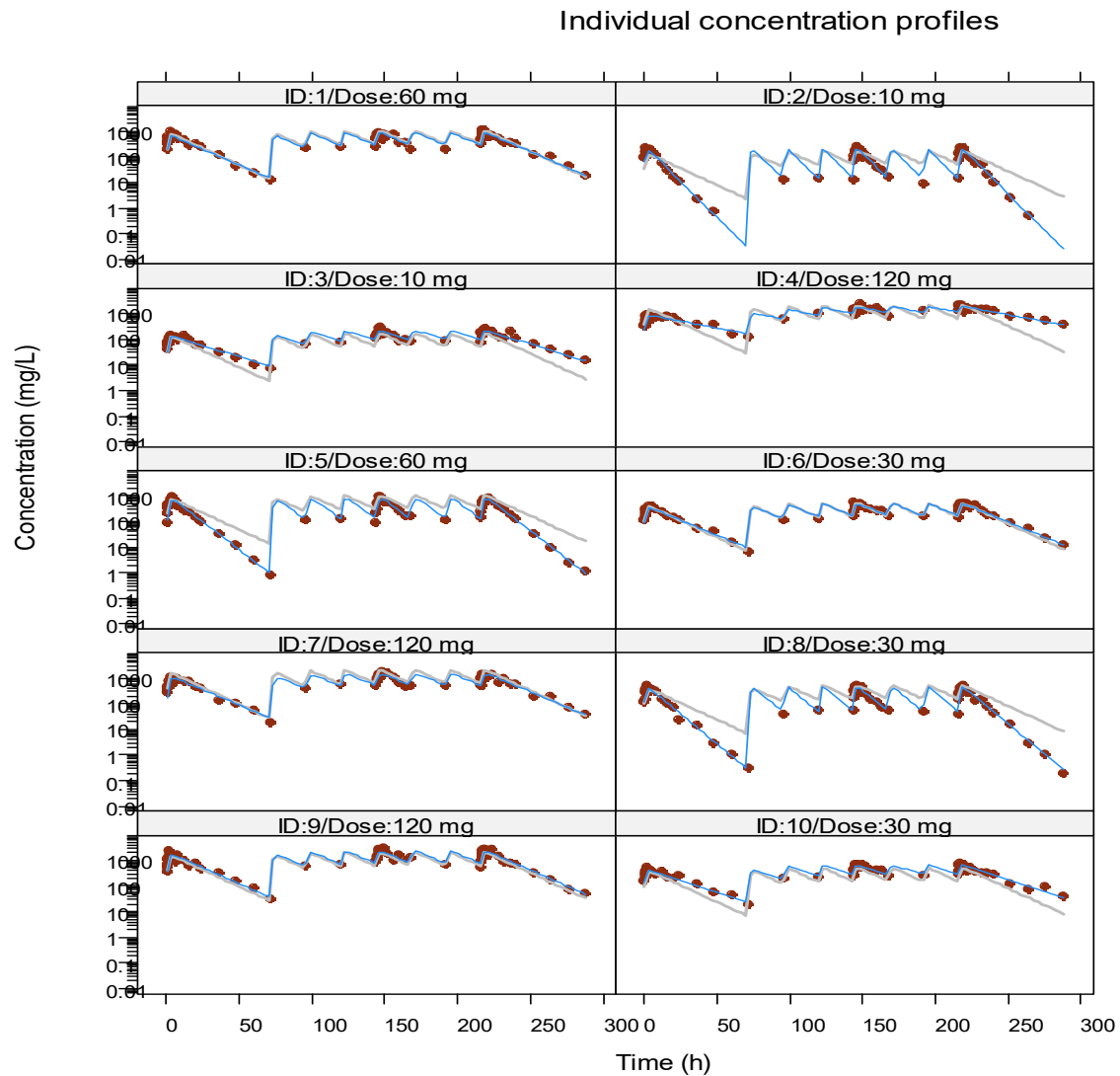
Results can be exported to pdf or html

The screenshot displays the ShinyMixR web application interface. On the left is a dark sidebar with navigation options: Model overview, Widgets, Edit model(s), Run model(s), Parameter estimates, Goodness of fit, Fit plots, Analysis results, and Settings. The main content area features a 'Show results' button and two radio buttons for 'PDF' (selected) and 'HTML'. Below these are two panels: 'Model(s)' containing a list of analysis runs (run1, run2, run3) with run1 highlighted, and 'Result(s)' containing a list of generated files: GOF.pdf, GOFst.pdf, IndFit.pdf, and ParTable.pdf. The nlmixr logo is visible in the top right corner of the interface.

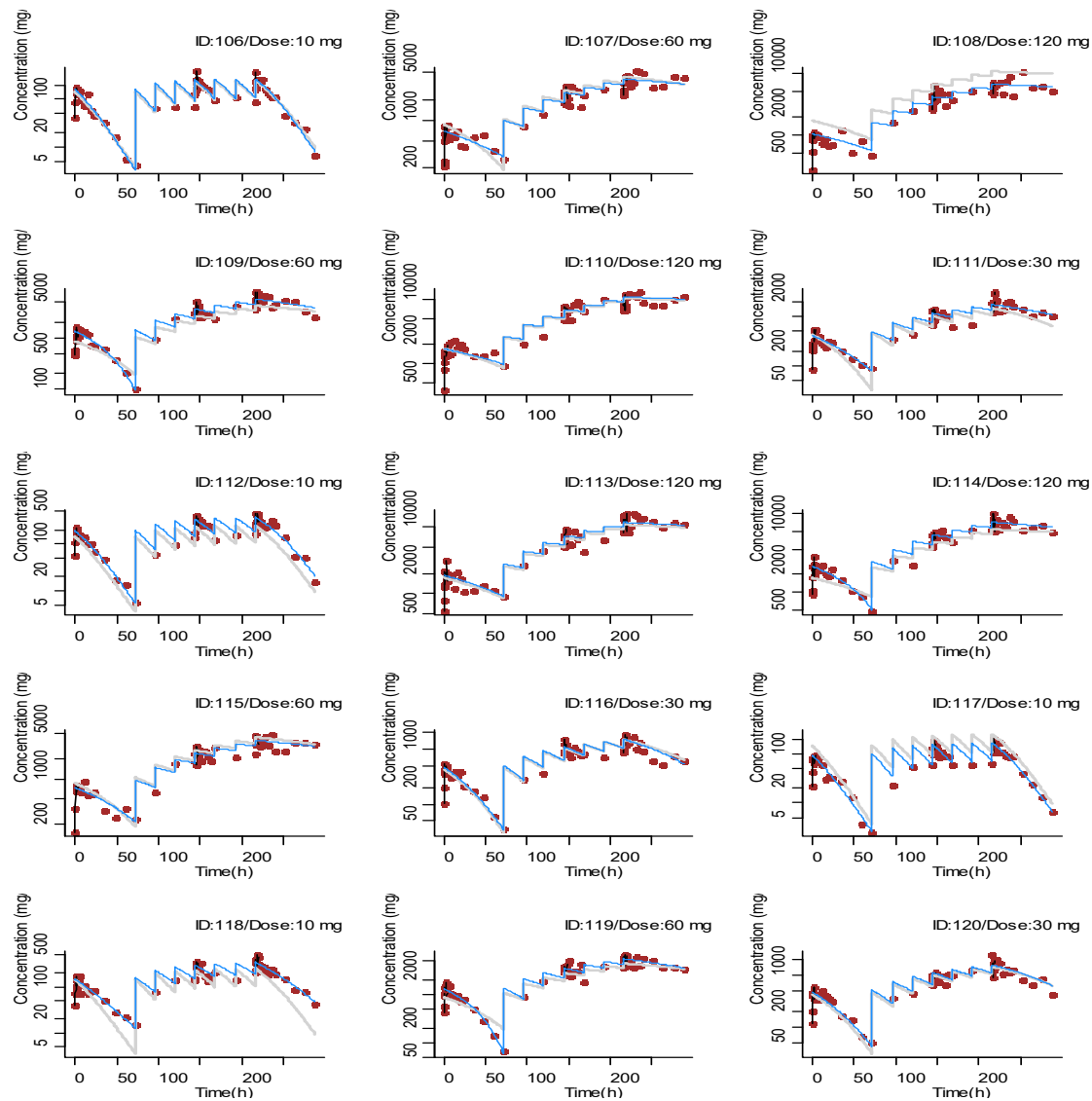
nlmixr performance

- 4 different dose levels (10, 30, 60 and 120 mg) of 30 subjects each as
 - single dose (over 72h)
 - multiple dose (4 daily doses)
 - single and multiple dose combined
 - and steady state dosing
- Range of test models:
 - 1- and 2-compartment disposition
 - with and without 1st order absorption
 - linear or Michaelis-Menten (MM) clearance
- A total of 42 test cases
 - all IIVs were set at 30%, residual error at 20%
 - overlapping PK parameters were the same for all models
- nlmixr estimation routines compared to NONMEM FOCE-I

Example full profiles (linear elimination)



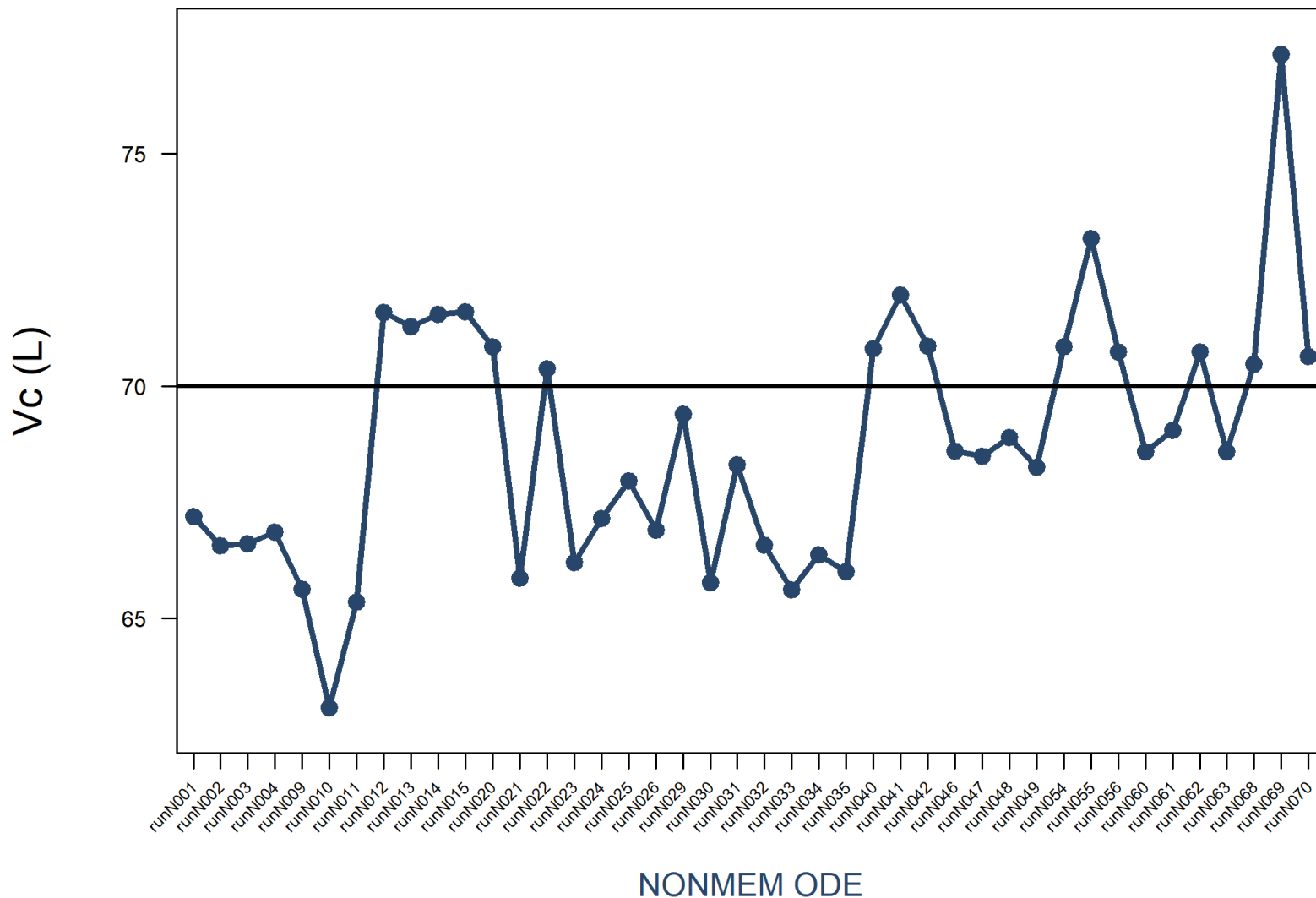
Example full profiles (MM elimination)



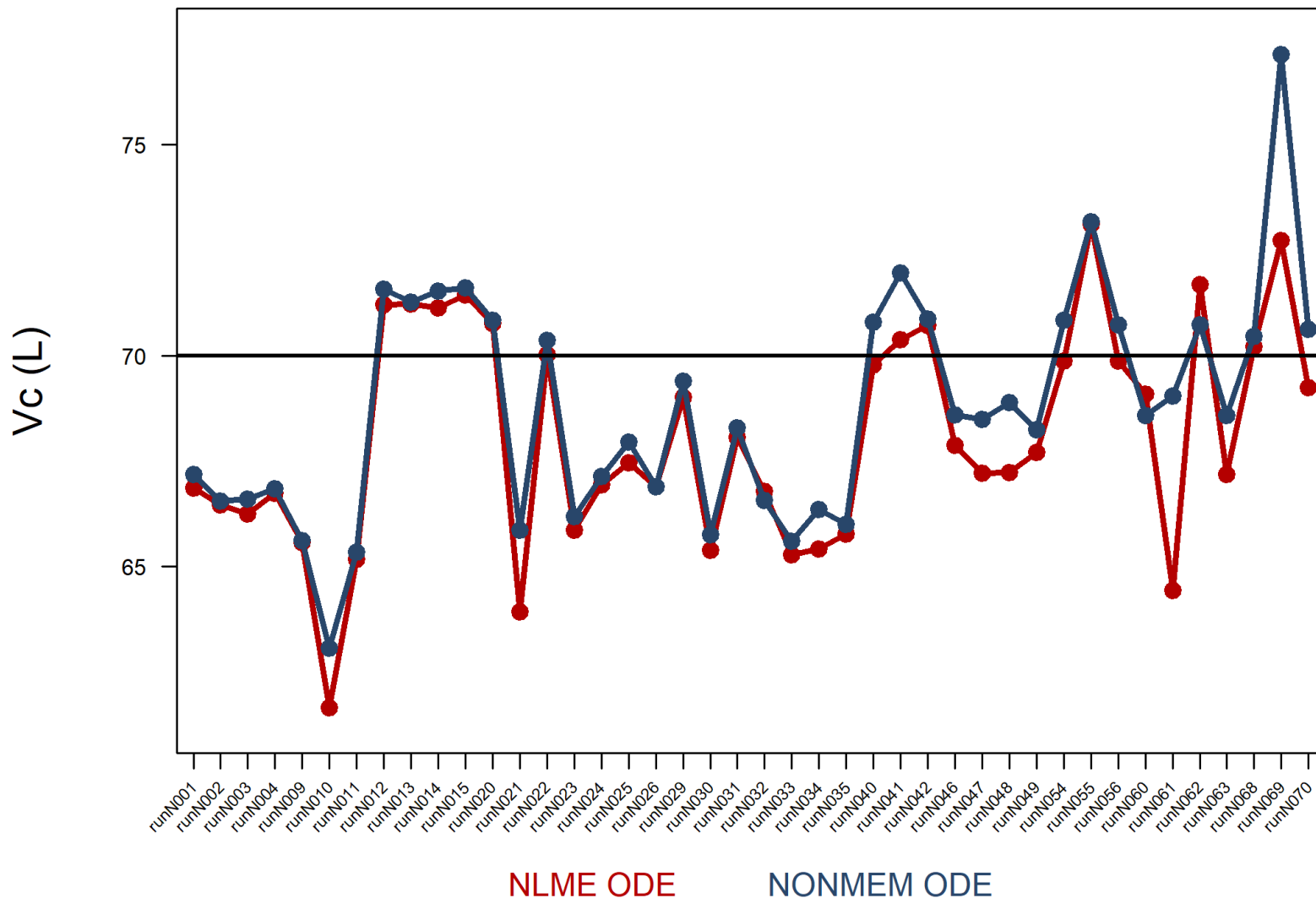
Vc is available in all models:

Theta estimates using NONMEM FOCE-I and ODE implementation

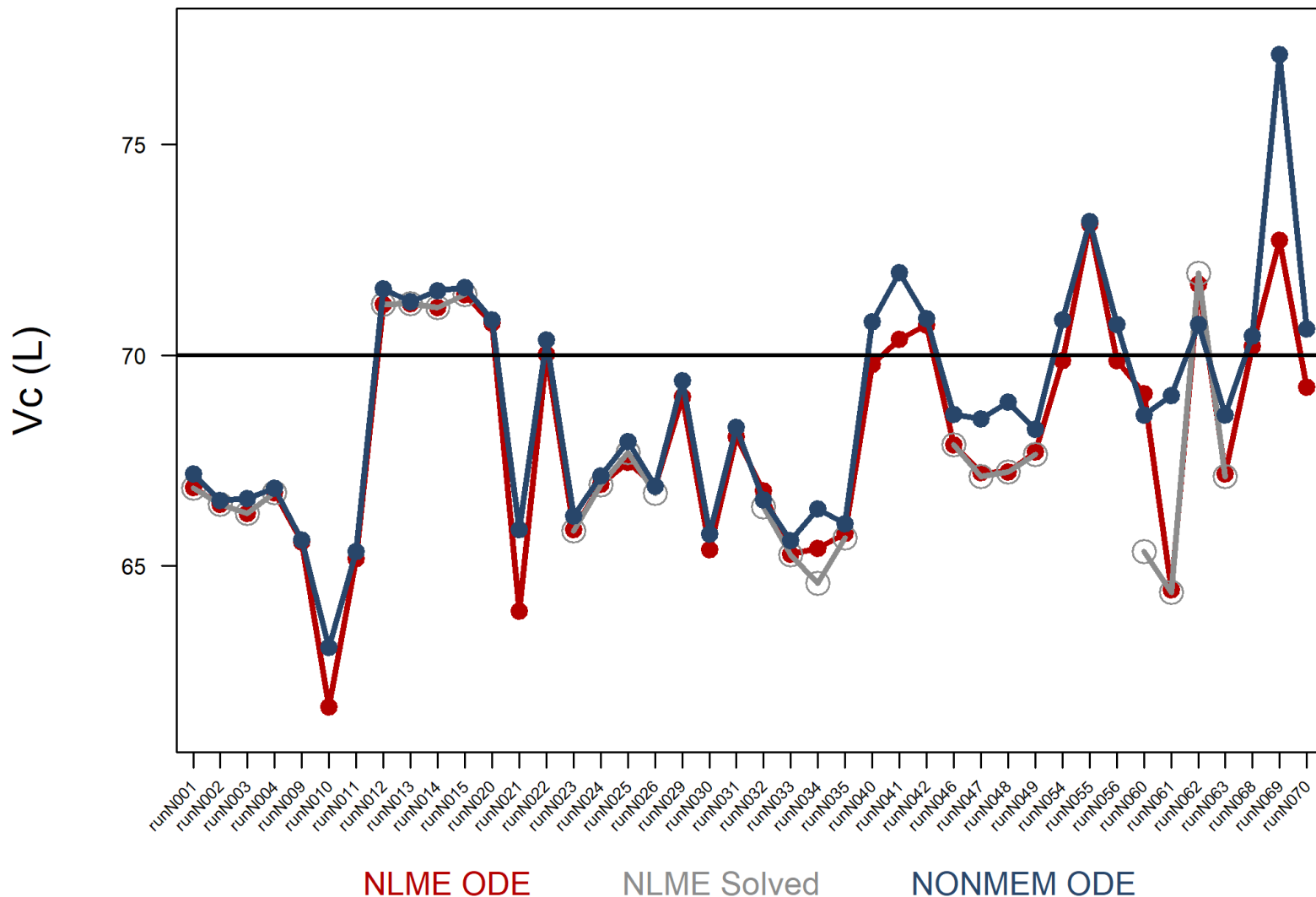
Horizontal black line: value used for simulation



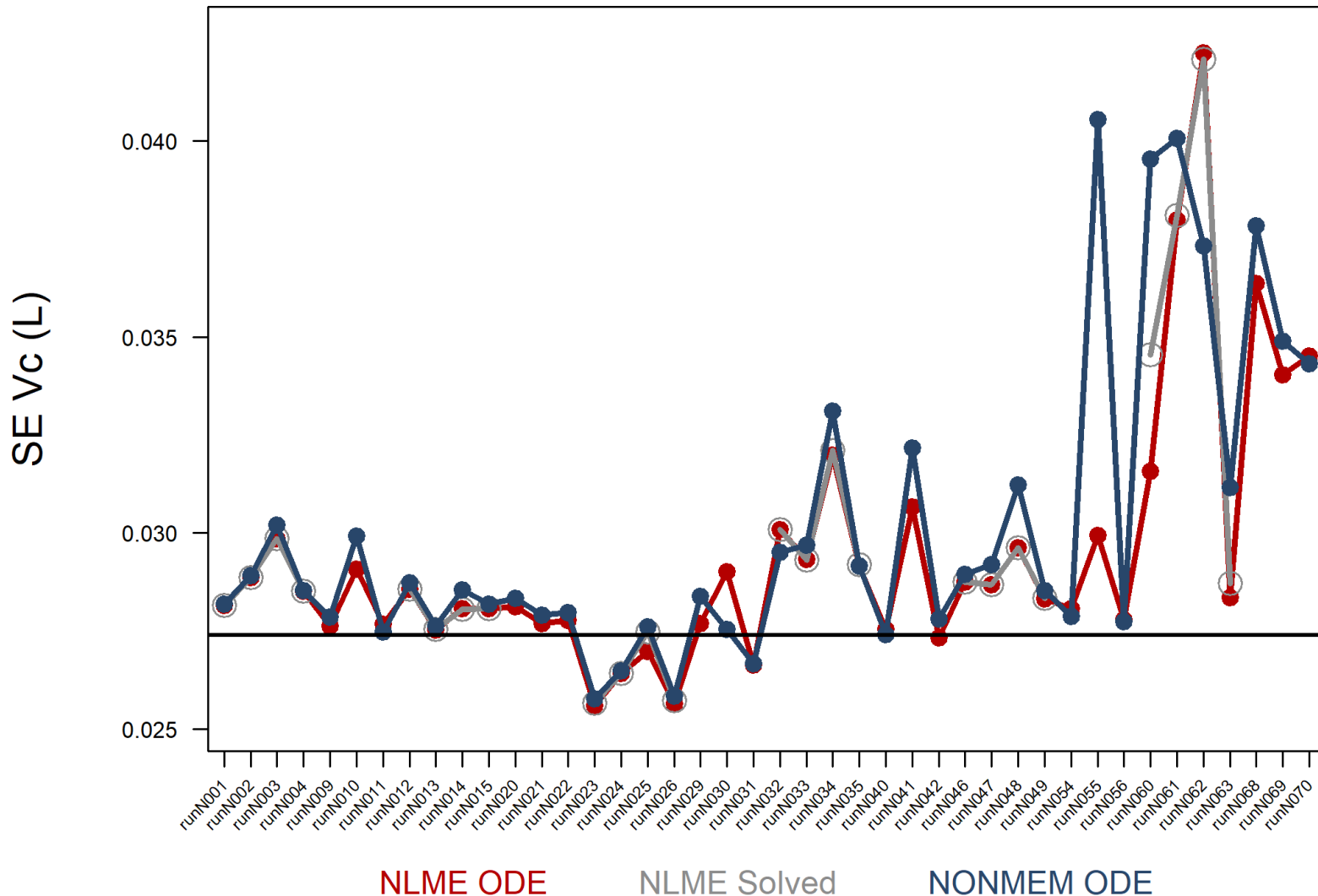
Red line: nlmixr/nlme estimates using ODEs



Non MM models also implemented using closed-form solutions: Grey line: nlmixr/nlme estimates using closed-form solutions

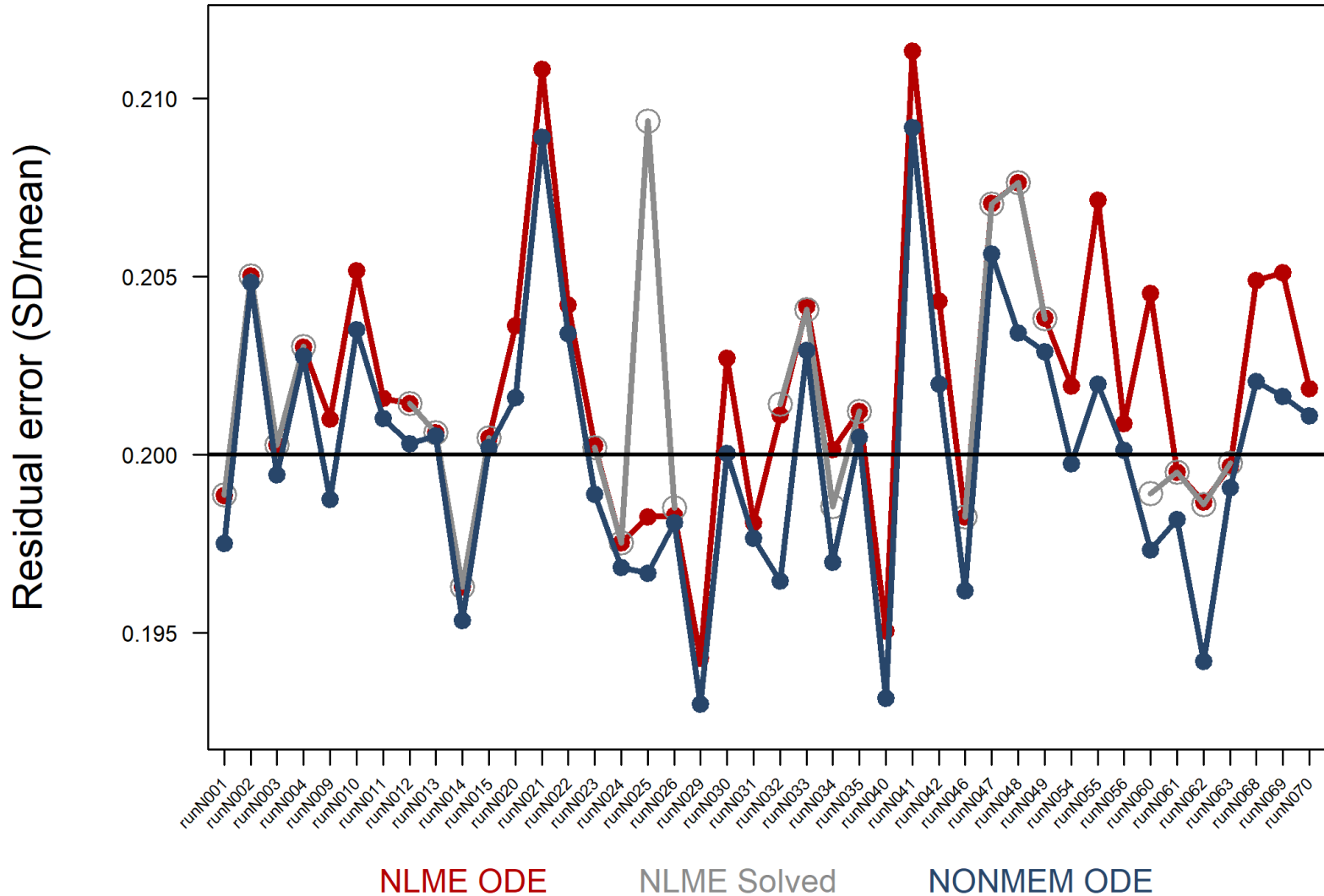


SE of theta estimates for Vc are very comparable

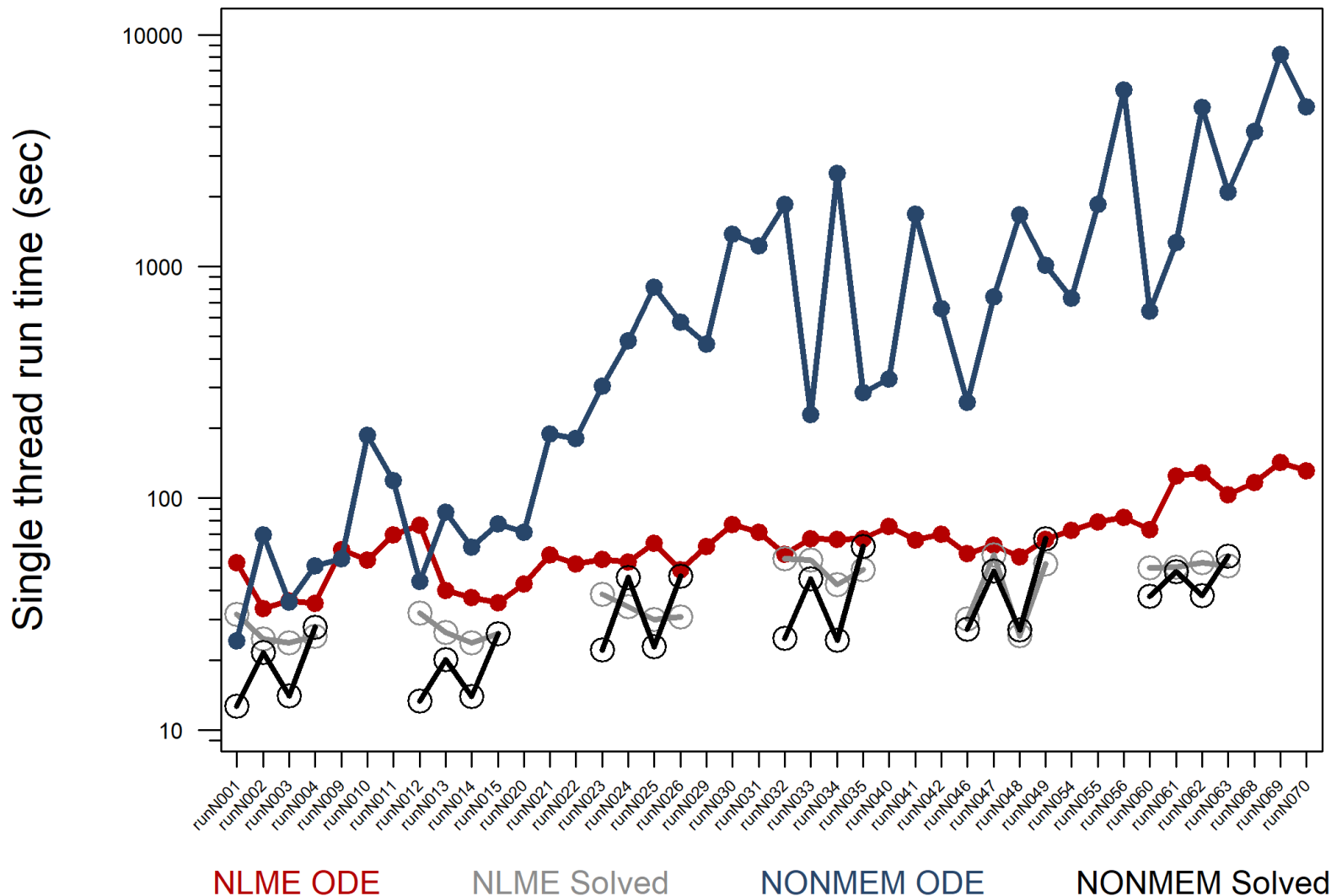


Residual error is well-estimated

Horizontal black line: value used for simulation

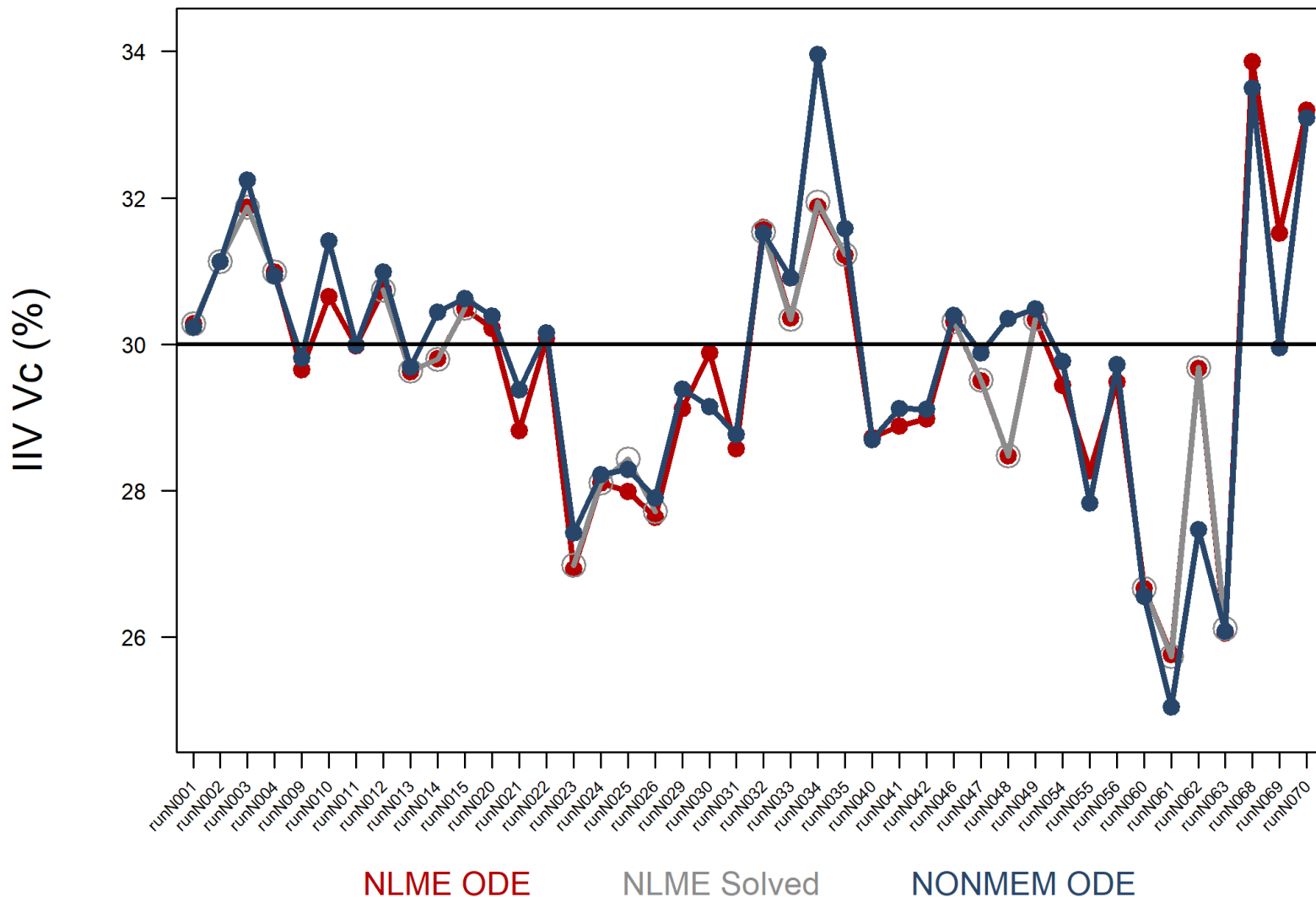


Run times are perfectly acceptable, and often lower than NONMEM ...but currently only single-threaded...

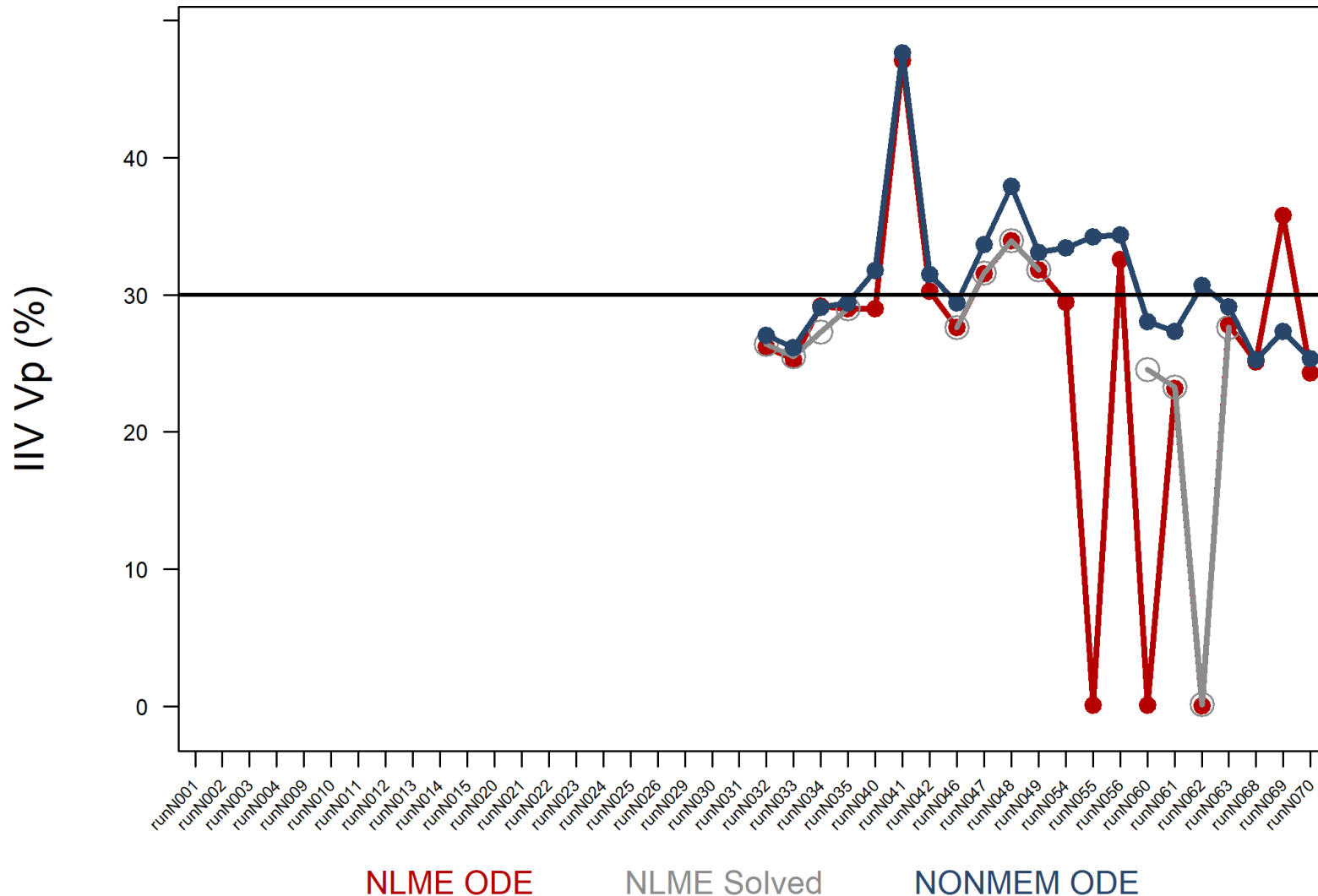


For Vc, Omega (IIV) estimates are also very comparable

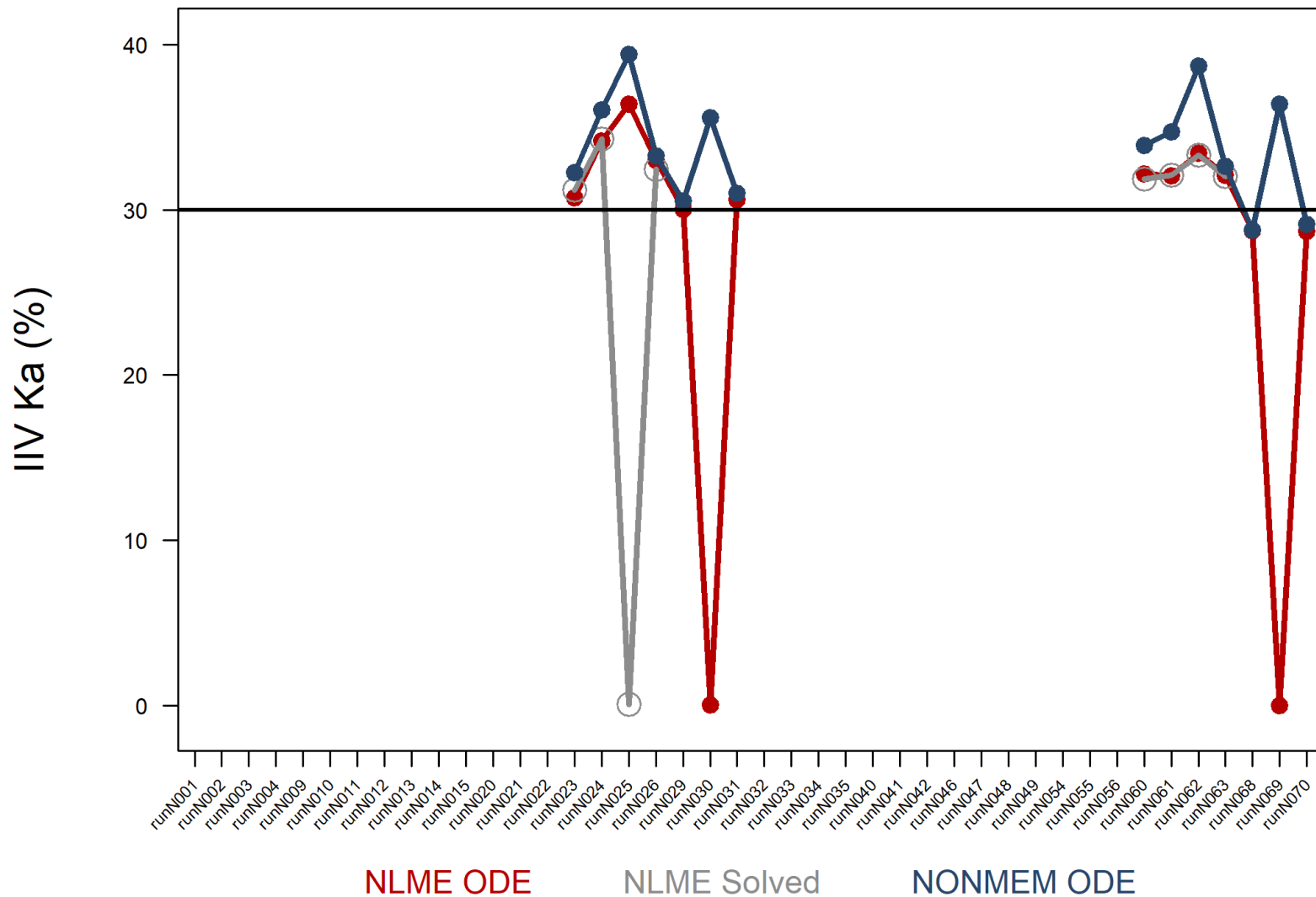
Horizontal black line: value used for simulation



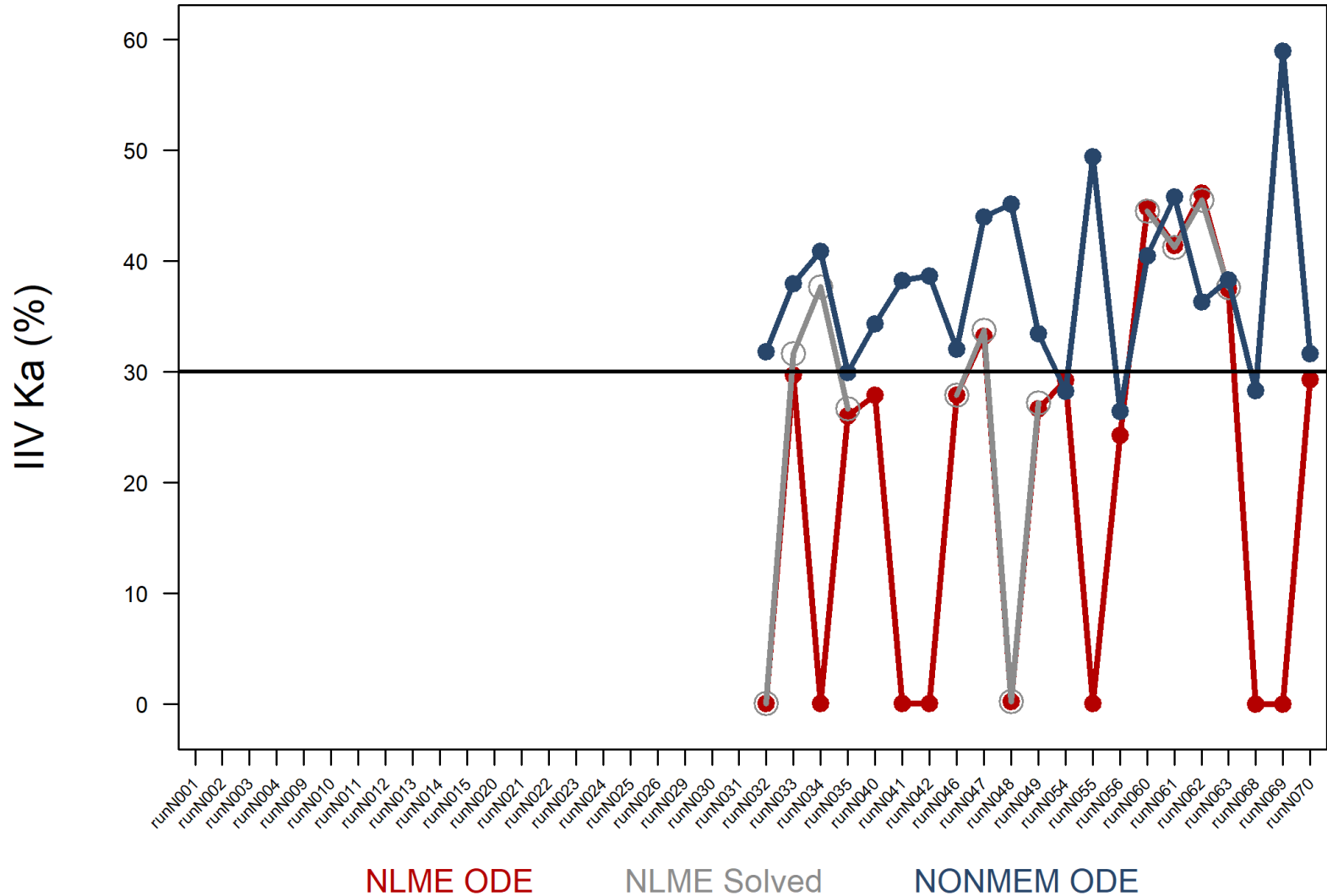
But if we examine V_p ...
the IIVs are often estimated close to zero



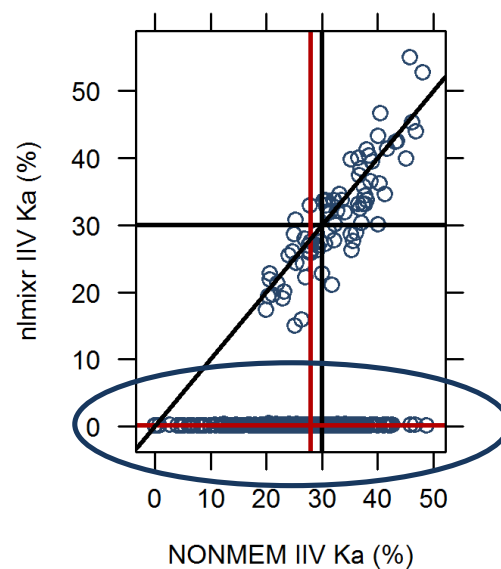
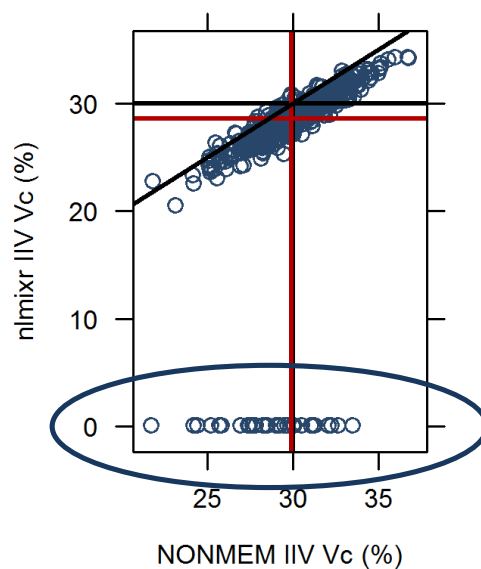
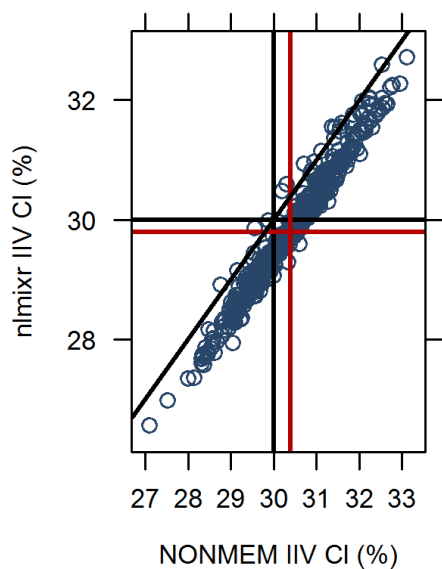
...same with Ka...



...and Q



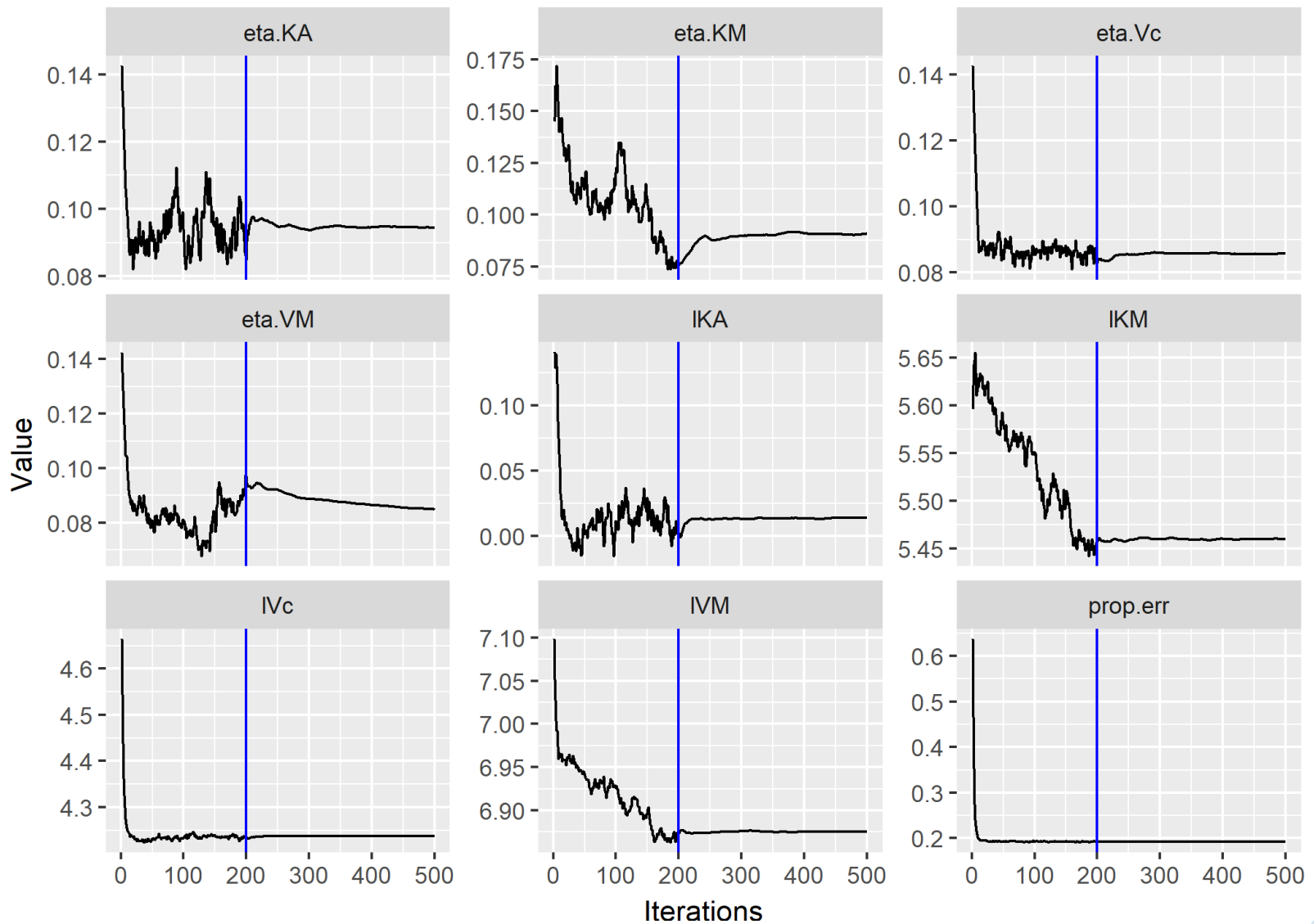
A large fraction of runs with IIV=0 for Ka for 500 sparse datasets: 91.1% for **nlmixr/nlme** vs. 2.2% for NONMEM FOCE-I



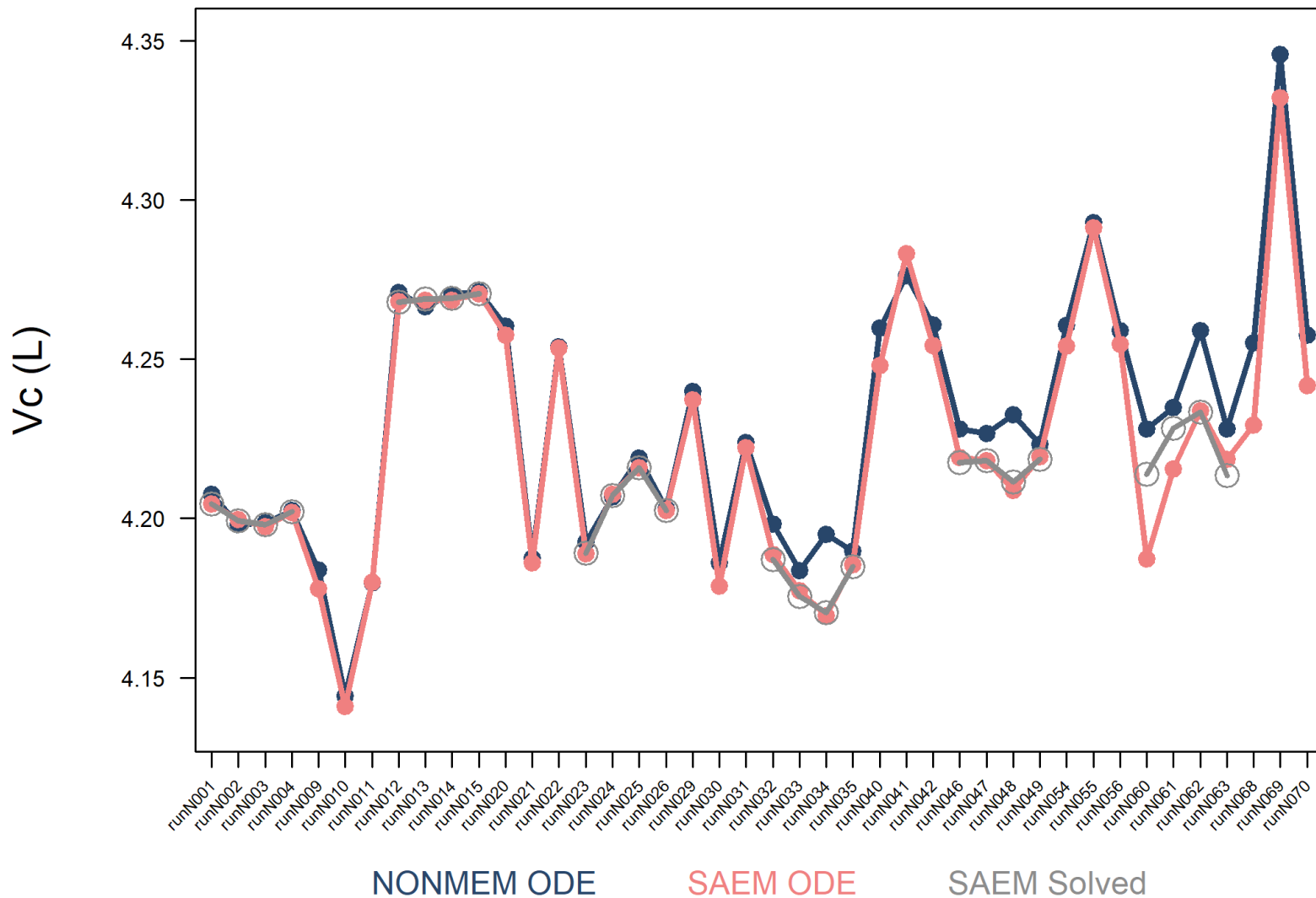
Disappointing results?

- Findings are in line with earlier experience with nlme
- Bob Bauer claims nlme is somewhere between ITS and FOCE (personal communication)
- However, nlme in `nlmixr` provides a gateway into nonlinear mixed effect modelling for statisticians...
- With the machinery in place, the groundwork is laid for other/better estimation routines, like SAEM or FOCE-I...
- SAEM currently also available in `nlmixr`: so how does SAEM perform?

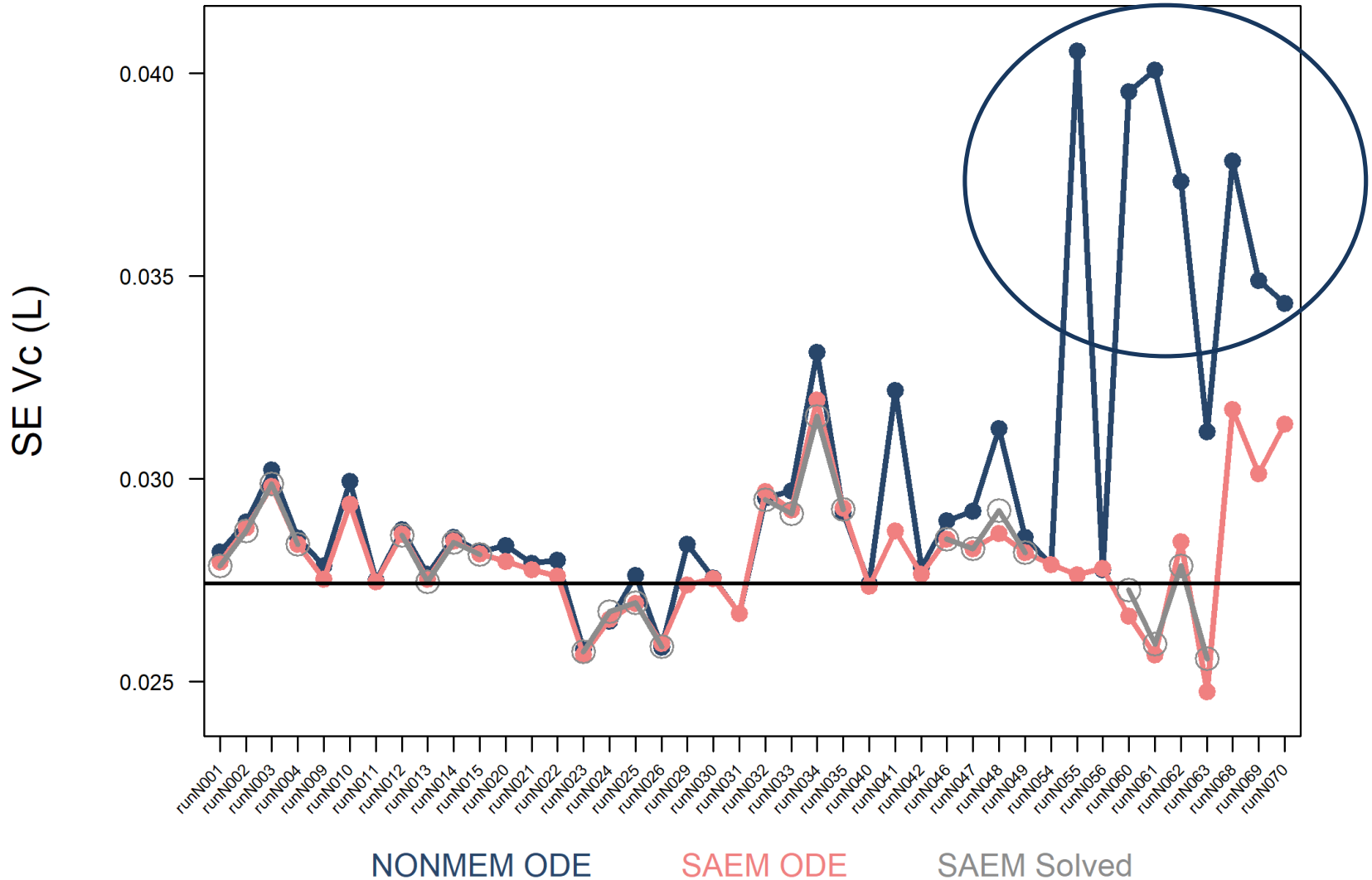
Traceplot for parameters from one of the `nlmixr`/SAEM models



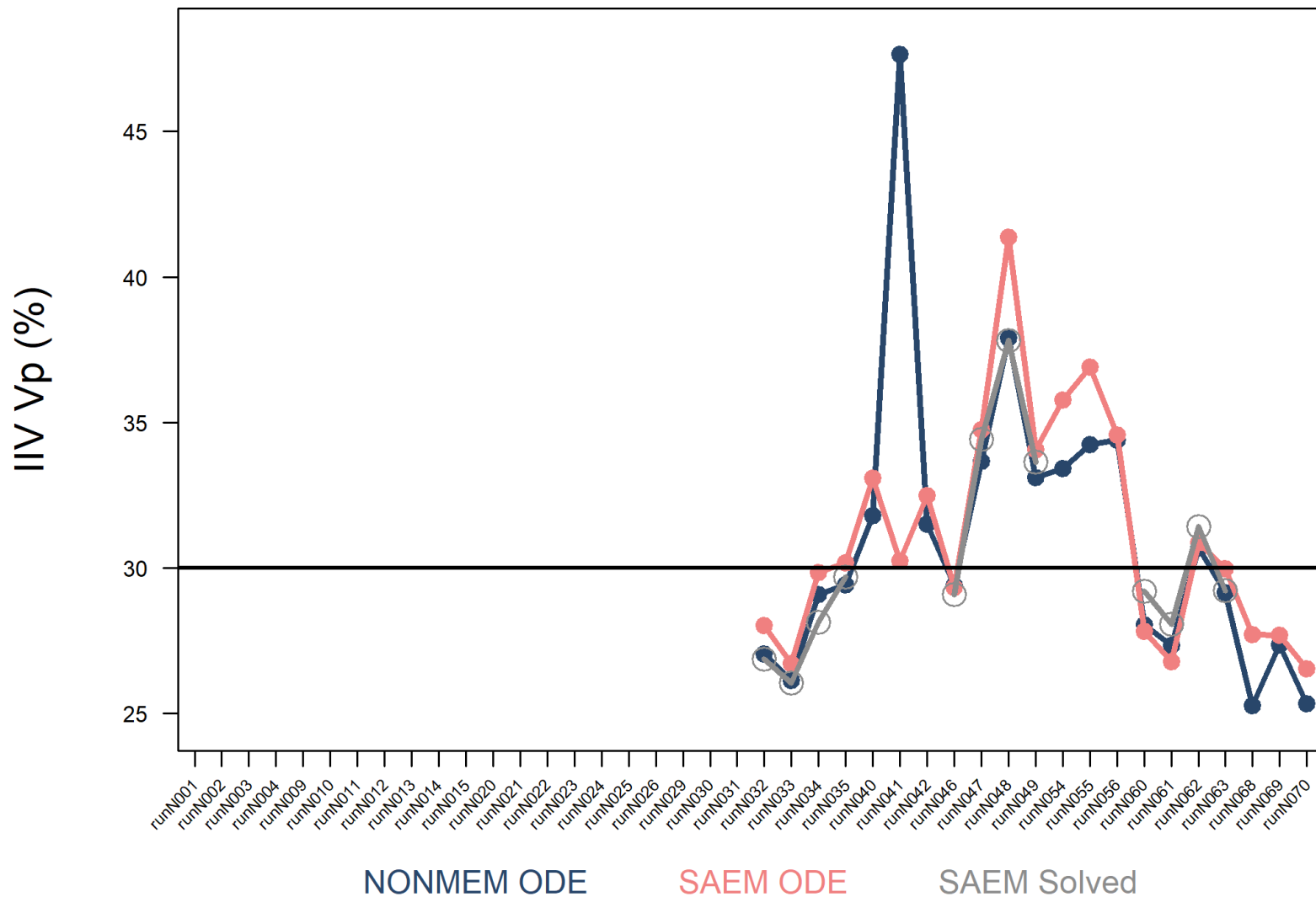
Thetas for **nlmixr/SAEM** for Vc behave very nicely compared to **NONMEM...**



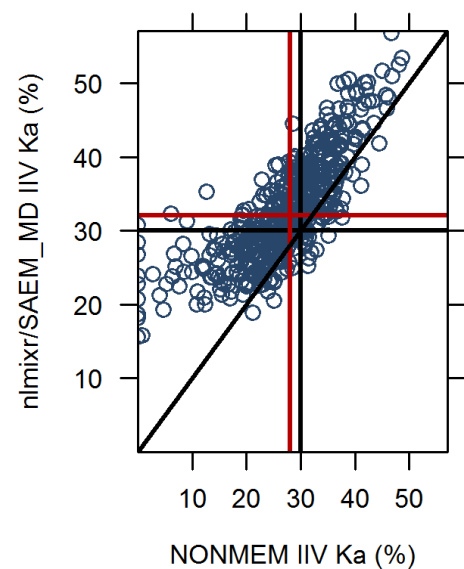
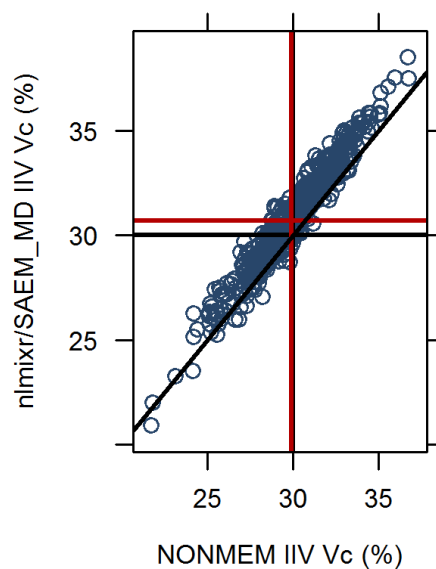
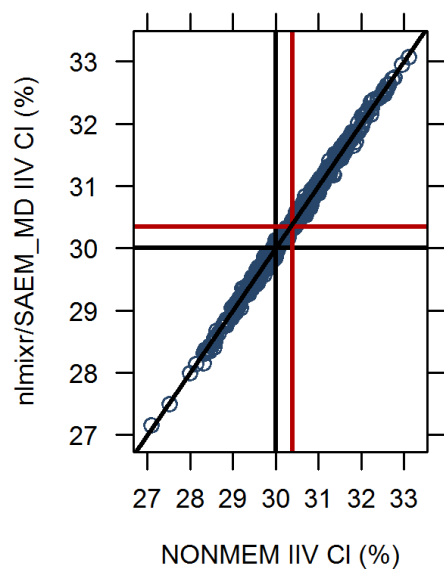
... and SEs for Vc seem to be even better estimated with nlmixr/SAEM than using NONMEM...



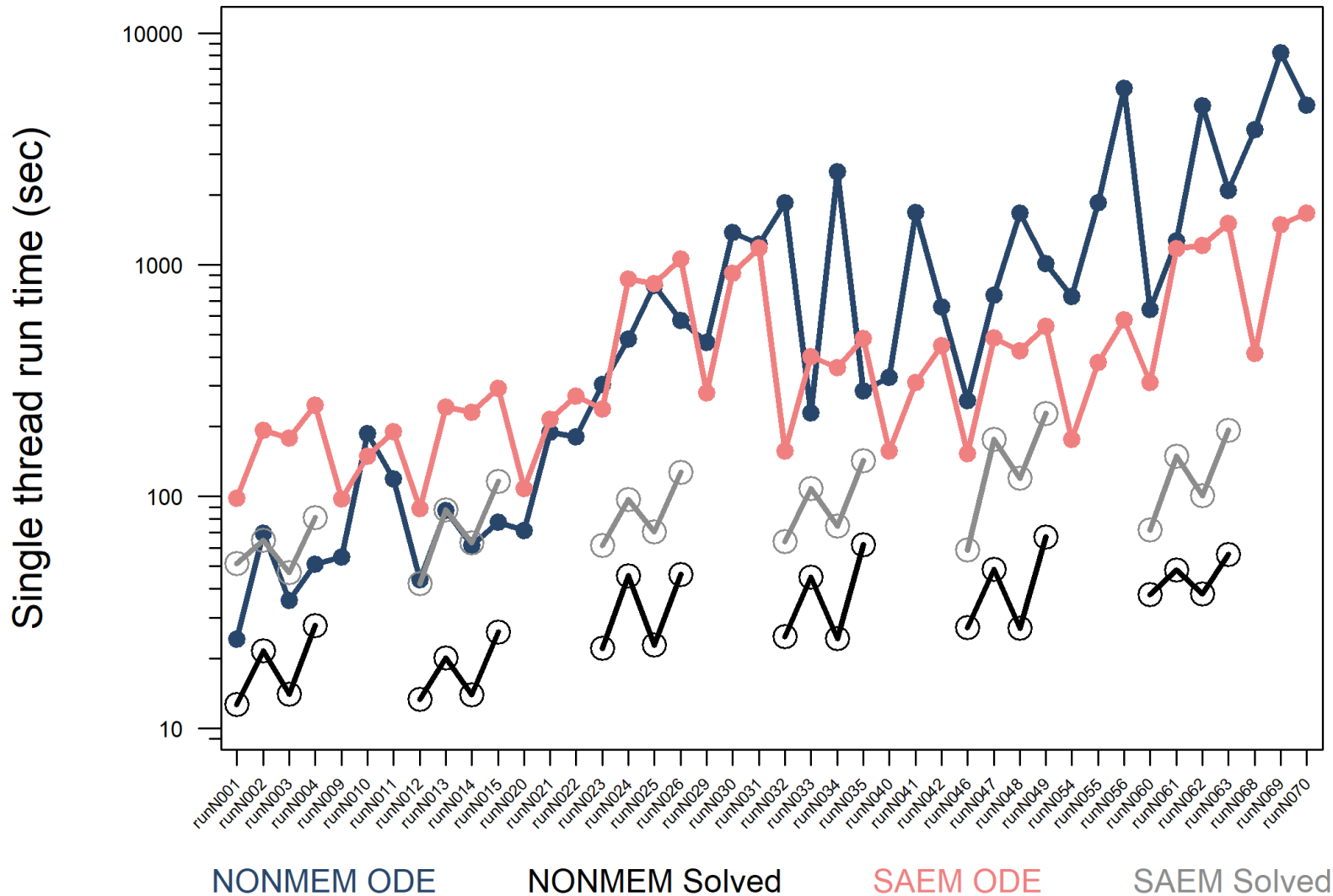
...and IIVs for **nlmixr/SAEM** for V_p show none of the close to zero behaviour observed with **nlme**



And no IIVs of zero with nlmixr/SAEM with sparse data



nlmixr/SAEM is slower than nlmixr/nlme but still workable



More good news?

- `nlmixr` also has an adaptive Gaussian quadrature algorithm (like NONMEM's Laplace and higher) allowing fancy models
- `nlmixr` also has single subject dynamic models e.g. for complex system simulation and estimation (mcmc algorithm)
- Steps to implement ordered categorical models and count models in the SAEM algorithm
- Elementary implementation of VPC and bootstrap functionality
- Serious progress into multi-threaded simulation that will lead to multi-threaded estimation
- Implementation of FOCE-I under construction

What's next?

- We need you!
- Field-testing: real-life examples
- Improving computational efficiency of estimation algorithms (e.g. within-problem parallelisation)
- Error-trapping
- New features implementation
- Etc, etc...
- This presentation will be made available on the bookdown site nlmixr.github.io